

QuantPrime

- a flexible tool for reliable
high-throughput primer design
for quantitative PCR

<http://www.quantprime.de>

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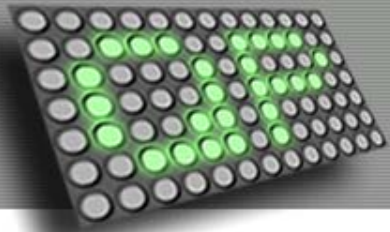
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Samuel Arvidsson, March 11th 2009, qPCR 2009 Meeting Freising, Weihenstephan

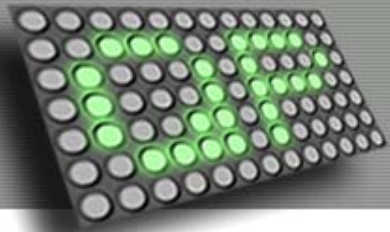
Reference:

Samuel Arvidsson, Mirosław Kwaśniewski, Diego Mauricio Riaño-Pachón and Bernd Mueller-Roeber: QuantPrime - a flexible tool for reliable high-throughput primer design for quantitative PCR. *BMC Bioinformatics* 2008, **9**:465



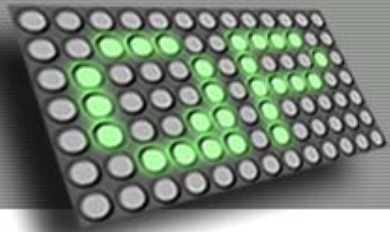
Background

- Projects going on in our group
 - Leaf growth and senescence (mainly *Arabidopsis*)
 - Salt stress in rice
 - Systems biology of biomass accumulation (*Chlamydomonas*)
- Working with transcriptomics in plants and algae
 - Transcription factors (TFs)
 - Membrane channels, pumps and transporters
 - Expressed on low levels → microarrays not feasible
- qPCR “platforms” for different gene groups
 - Covering many transcripts (hundreds to thousands)
 - Keep costs down:
 - SYBR Green I based assays
 - Small reaction volumes



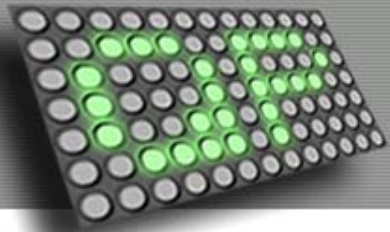
Why develop QuantPrime?

- Need of software for creation of qPCR platforms for high-throughput transcriptomics
- Loads of primer design software available, but...
 - there are important features missing or not suited for high-throughput design (most free software)
 - they are very expensive and not customizable for our own demands (most commercial software)
- Companies are offering such services, but...
 - on a fundamental research budget quite expensive and the service often a “black box”



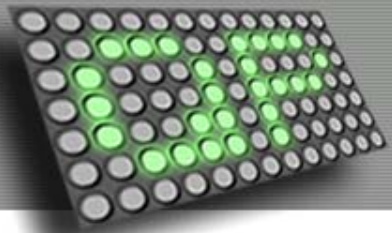
What we need...

- Automated design which can...
 - directly use common annotations of sequence sets
 - have extensive basic primer parameters (melting temperature, GC content etc.) with ranking of primers
 - organize sequences, transcript identifiers and primer pairs well
- Automated specificity checking which should...
 - do checking against transcriptomes and genomes
 - score the specificity of the primer pairs
 - handle splice variants of transcripts
- And of course, it should be reliable and fast...



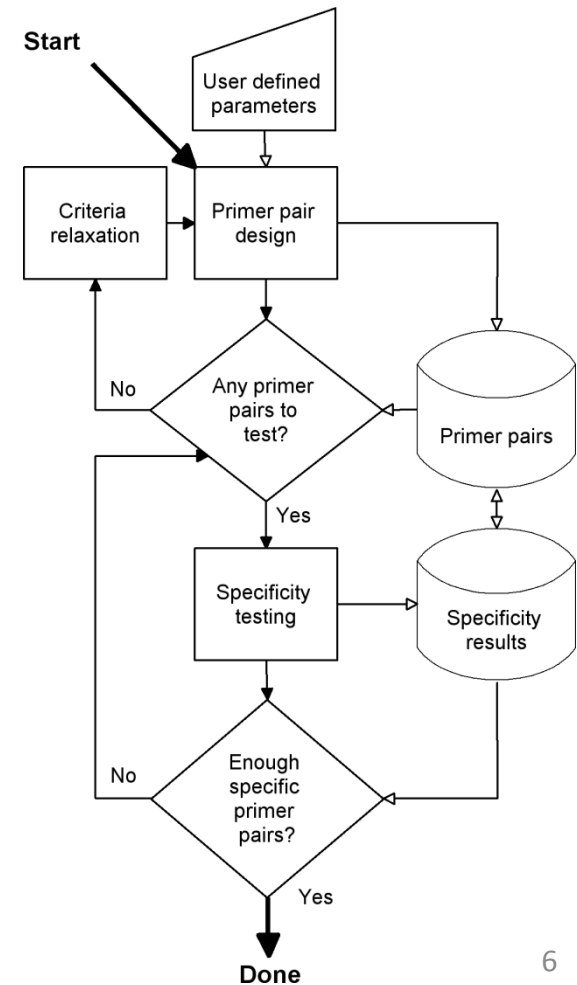
So...

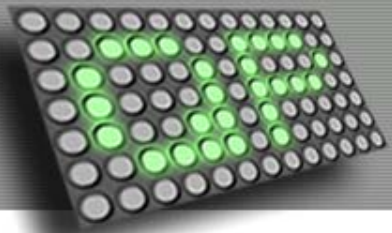
- I needed about 200 primer pairs for my project
- Instead of ...
 - paying a company for the primer design (too easy)
 - spending the time checking and testing the primer pairs designed by simple tools (too much manual work for an engineer)
- ... I decided to invest time in making a tool that does just what I needed (more fun)
- After many discussions, much programming, more GUI design and some extensive testing that tool ended up becoming “QuantPrime”



What can QuantPrime do?

- Designs primer pairs and hybridization probes for expression measurements with qPCR
- Organizes...
 - parameters for design and specificity testing
 - sequences and annotations
 - primer design projects
 - primer pairs
- Predicts specificity of primer pairs



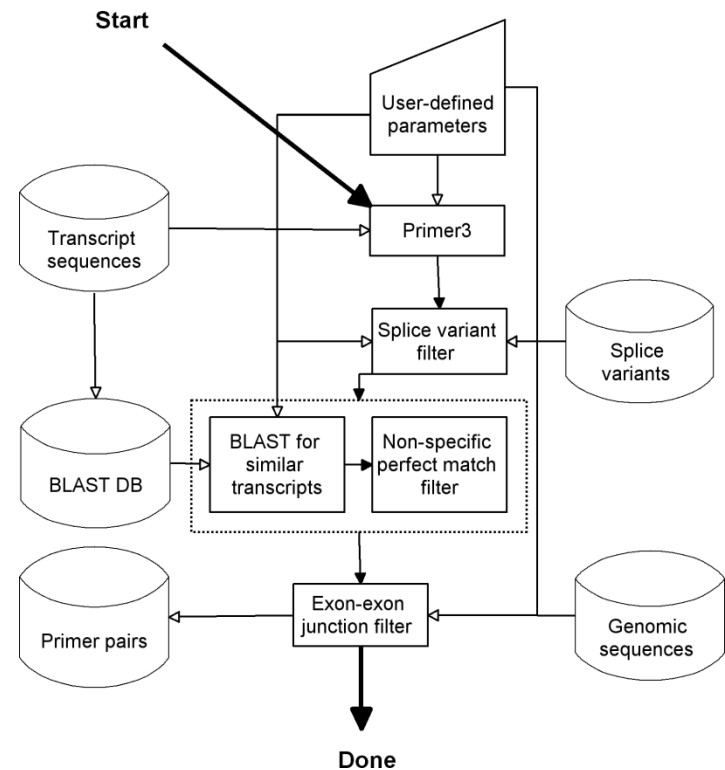


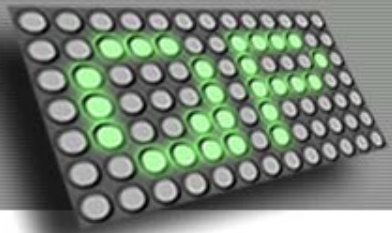
Automated primer design

- Primer3 with extra parameters
 - Primer 3'-end G/C-content limitations
 - Transcript 3'-biased design, when using oligo-dT cDNA synthesis
 - Filtering of primers against similar transcripts
- Genomic DNA amplicon avoidance
 - Exon-junction spanning primers
 - Large-intron-spanning
 - No manual annotation necessary

GGTGGACATTCTCTGTCTCCTCGAGTCAACAAGG*ACGAGTGGGTGGTGTGTGCATTTTTTCAAAGAATTCTGGAGGGA
AGGGAAGTTTTTTGTTCTCGGATTCCATGCATGCGCAGCCATGCGGTGATGTA**CCAGATTGAGGACACRAGGTCGT**CTTTG

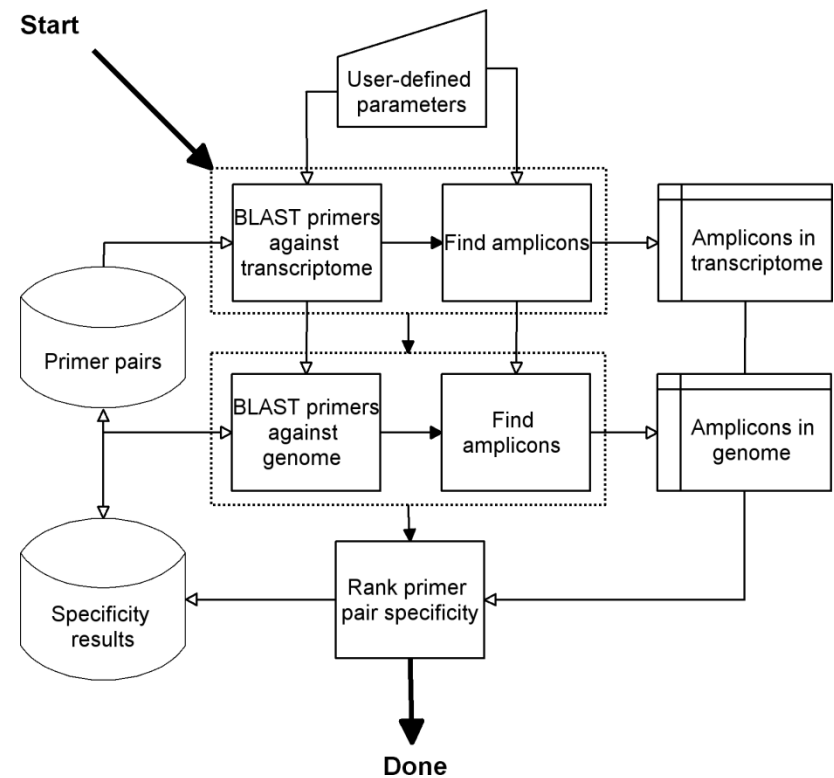
- Hybridization probes
 - Together with primer pairs or alone (e.g. for QISH)

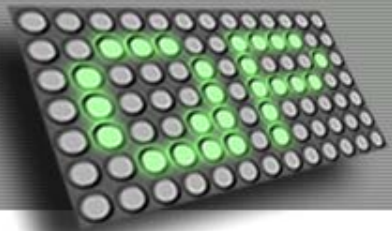




Specificity testing

- Amplicon finder based on BLAST used on
 - transcript sequences
 - gene model sequences
 - unannotated genome regions
- Ranking of primer pair specificity





Public web server

- Integrated graphical user interface

Primer pair information

Transcript identifier: AT1G76130.1 - ATAMY2, AMY2, AMY2/ATAMY2 (ALPHA-AMYLASE) alpha-amylase, ch11:28566175-28568970 FORWARD

Forward primer
Sequence: ACTTACTCATCCGGCATTCCC (22 b)
Melting temperature: 62.5 °C
G/C content: 54.5 %

Reverse primer
Sequence: TTGTCCCTCCTAATGTCAATCAG (reverse complement: CTGATTGCATTAGGAGCGCAA)
Melting temperature: 61.3 °C
G/C content: 45.8 %

Amplicon
Size: 106 b
Melting temperature: 61.1 °C
G/C content: 43.4 %
Optimal annealing temperature: 60.3 °C

Alignment with transcript sequence
GGTGGATTAGCTTCTCGTTTCCTGGTAATCGTCTGTTTTCGGTGAATCTGTTGGAGTTTCATGGAATCTTG
TCTGGACATGGCTACTATAACAATCTTTTGTAGTACTGACGACCAAACTGATTCGGTGGTTATACGGATGGAA
GGGAAGTCAITTCACAGCATATAATGGGAATCCATAAATATGATTTGGTGGAGAACTTGGATGGTAAAGTCTTGAC
ATCCGGAATCCGGGTTTACTTTCGATGTTGGCAACACCATTCTGCTGCTGACCGGAAGTTTATTCACAGGA
CCTTTATCTACTAACTCAGCATATGGCTGTGGACATCTATTGAAATCCTTACCTGCGCAAGATGAAACAATAACAAGTA
GAGCTATGGCTGATATGATCATACTATCGTGTGGGCACTAGAGGACATGGTGGAAATGATAAACCGTTATGATGGA
ATTTCAATCCGTTGGATGACACCGCTGACTTCTGTACCGGAGGACTGGTAAACGAGACAGAGGGATATTTCAA
TGGAGTCCAAATGTTGATCAGCACTACGATTTTGTAGGAAAGATATAATGGATGGCTCTTGTGTTACGACACCG
TGGGTTTCAAGATTTCTGTTACTTTCAGGGTTATTCAGCAACTATGTAAGGAATACTTGGCCGACGGA
CCATTAATCTTGGTTGGAGAAATGTTGGAAATTTGCAATCAATGGCATTGGCTAGACTAATAACAATAGCCATAG
ACAGCGTATAATCACTGGATCGTGGCAAGGGACAGATCTCTGCTGATTGACTTCACAAATAAAGGAATCTGGCAGG
AAGCCGTAAGGGCTCAGTATGGCTTATGATGGATGCCAAGGGAAGCACCGGGTTAATGGATGGCTTGGCCTCAAGA
GCTGTCACATTCCTGATAACCATGACACTGGCTTACTACGGCTCATTGGCCGTTCCCTCACACAGCTTATGGAGGG
CTATGATATACTTCTCTCTCCCGCATTTCCTGTTGTTCTAGCATCTTTACGATTTGGGAGGCTCAAGATG
ATGAGATTGTAAGT GATGGCATTTGGGGGGGCGAGATTCACAGTAAAGTCACTGTCCGTTTTTAAAGAGAG
ATCACTTATATGGACGATTTGTTGGAGAAATATGATGAGCTTGGACGGCTCTTGGTCCCTTGGTAGAG
ACTGGACTTAGCAACAGTGGCCATCGTATGGCTGGCAAGATTAATCTTTGTCTATCTAGCTTTGAGTCCA
AAGAAAATATATGTAATCTTCTGCTACCGGTTTACTATAAGTAACTTTTGTCTATCTAGCTTTGAGTCC
TATAAGTTATTTAATGATGAAATTTGATCTTTCGTGGAATGTTTGG

Specificity test results
Overall score: 100%
cDNA specificity: Good
Single primer specificity: Good
Amplifies genomic DNA: No

Results

Favour primer pairs
 not amplifying genomic DNA (default)
 with high single primer specificity (when genomic contamination is not a problem)

Select best Select all Clear selection

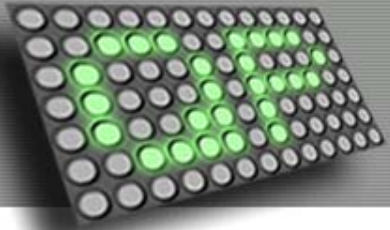
Fw sequence	Rev sequence	Amplicon size	Spans exon border	Rank score	Test results
AT1G01010.1					
ACGATCCTTGGAACTTGGCCTTC	AAACCGTGTCTCGCTCTGTCTG	114	Yes*	1.5084	✓
ACGATCCTTGGAACTTGGCCTTC	ACCAGAAACCGTGTCTGTCTG	119	Yes*	1.7827	✓
TACGATCCTTGGAACTTGGCCTTC	AAACCGTGTCTCGCTCTGTCTG	115	Yes*	2.5546	✓
TACGATCCTTGGAACTTGGCCTTC	ACCAGAAACCGTGTCTGTCTG	120	Yes*	2.89	✓
ACGATCCTTGGAACTTGGCCTTC	TTGTCTCGCTCTGTCTGTCTG	118	Yes*	2.4872	✓
CGATCCTTGGAACTTGGCCTTC	AAACCGTGTCTCGCTCTGTCTG	113	Yes*	1.6248	✓
CGATCCTTGGAACTTGGCCTTC	ACCAGAAACCGTGTCTGTCTG	118	Yes*	1.9101	✓
GATCCTTGGAACTTGGCCTTC	AAACCGTGTCTCGCTCTGTCTG	112	Yes*	1.9199	✓
GATCCTTGGAACTTGGCCTTC	ACCAGAAACCGTGTCTGTCTG	117	Yes*	2.1952	✓
CGATCCTTGGAACTTGGCCTTC	TTGTCTCGCTCTGTCTGTCTG	107	Yes*	2.6128	✓
AT1G40101.1					
AGCTCCCTCAGGACATGCTTTC	CCTTGGGGAAACATGCAAGATC	80	No	2.5594	✗
AGGAAGTGGGCGTTATGCATGG	AGTGGTACTCCACAAGCTTCCAG	65	No	2.9151	✗
AGGAAGTGGGCGTTATGCATGG	TGAGGTGTAATCCACAAGCTTCC	67	No	2.9203	✗
AT2G39760.2					
TGAGTCAGAGTGGGAAGGGGAAC	CTTCAAGCGCCCGATCAAAGTG	60	No	4.3214	✓
TACCGGGTCAACTCTGCCACTTC	ATAGAGTCCAGCAGTCCCAAGAG	70	No	5.6579	✓
GCTTATGGATCAGAGTGGGAAGG	CTTCAAGCGCCCGATCAAAGTG	65	No	5.6821	✓

Export primer pairs

Data: Complete Brief

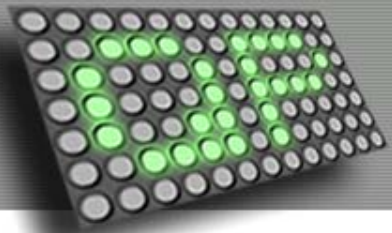
Format: European US Windows Unix Mac OS

- Currently 345 annotations of 295 species available
- Standard protocols for SYBR Green qPCR predefined, custom protocols possible



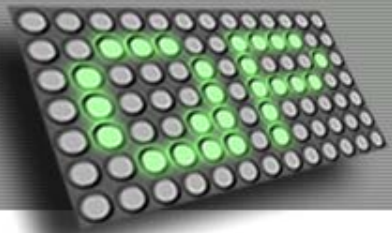
Tutorial...

- <http://www.quantprime.de/>



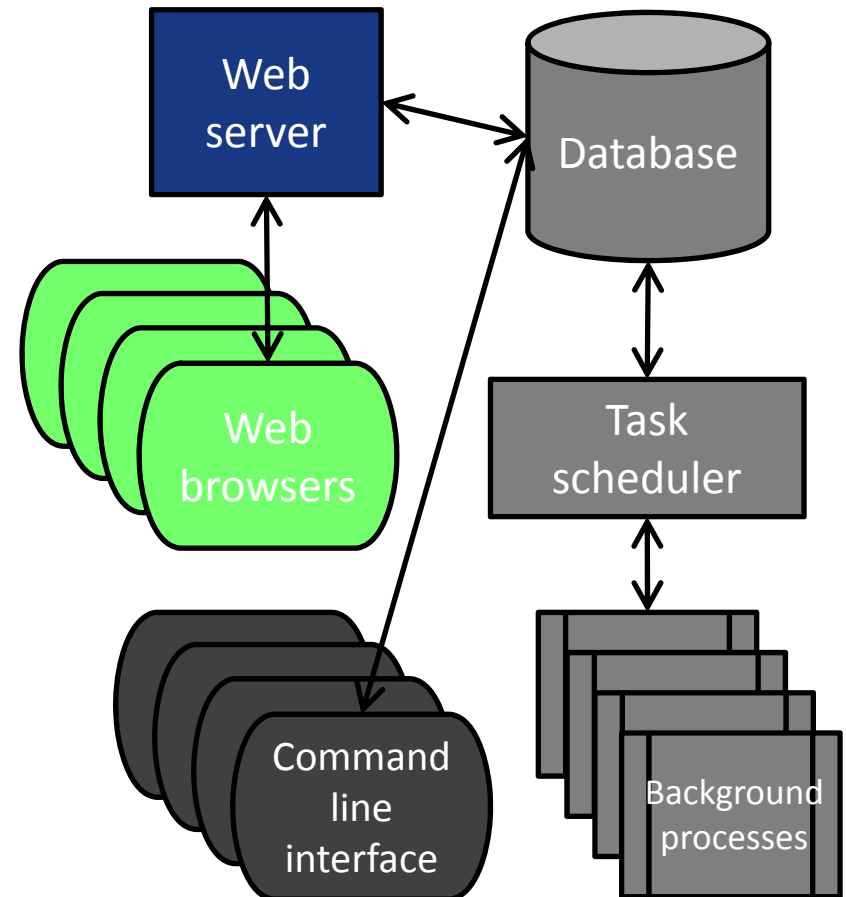
Local installation

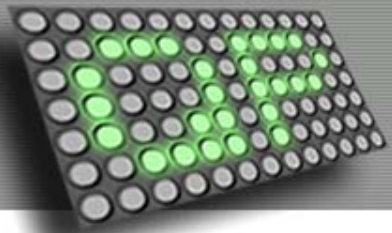
- Recommended for true high-throughput applications
- Calculation and database nodes can easily be installed on clusters
- Web interface for intranet applications
- Command line interface for integration in design pipelines



System architecture

- Easily extendable with new features and interfaces
- Possible to integrate in a design pipeline
- Suitable for parallelization



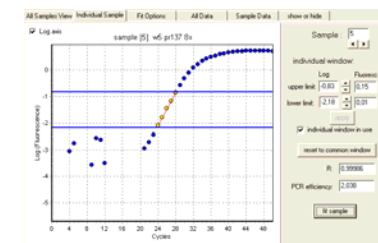
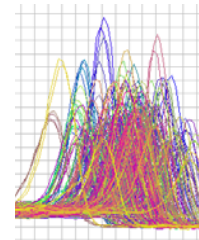
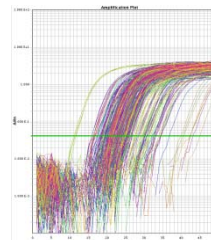
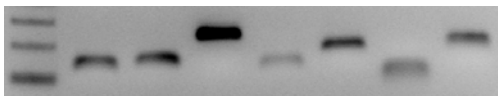


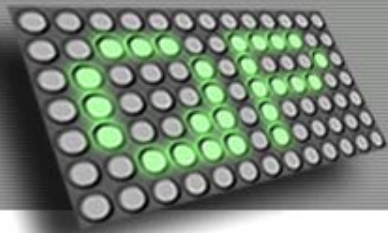
Experimental results

- Primer pairs designed by QuantPrime tested in real experiments

Organism	Total transcripts	Detectable	QC passed of detectable	Predicted gDNA-safe
<i>Arabidopsis thaliana</i>	175	165 (94 %)	160 (97 %)	149 (85 %)
<i>Chlamydomonas reinhardtii</i>	33	29 (88 %)	28 (97 %)	24 (73 %)
<i>Hordeum vulgare</i>	30	28 (93 %)	27 (96 %)	-
<i>Physcomitrella patens</i>	32	32 (100 %)	32 (100 %)	21 (66 %)
Total	270	254 (94 %)	247 (97 %)	194 (81 %)

- Quality control: amplicon size, melting curve, efficiency





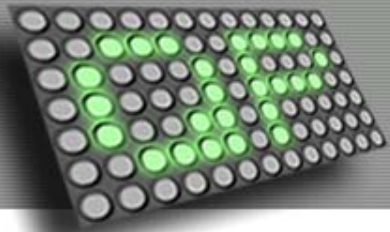
Performance results

Results of *in silico* benchmarking of QuantPrime

Species	Transcripts	Total search time	Average search time	Primer pair specificity ranking ¹		
				Acceptable ²	Good ³	Very good ⁴
<i>Arabidopsis thaliana</i>	5000	20:22:06	15 s	4916 (98%)	4323 (86%)	2534 (50%)
<i>Vitis vinifera</i>	5000	50:45:33	37 s	4765 (95%)	3927 (78%)	2315 (46%)
<i>Drosophila melanogaster</i>	5000	13:48:45	9.9 s	4894 (97%)	4075 (81%)	3096 (61%)
<i>Chlamydomonas reinhardtii</i>	5000	12:11:07	8.8 s	4568 (91%)	3999 (79%)	2349 (46%)
<i>Oryza sativa ssp japonica</i>	5000	83:31:12	60 s	4658 (93%)	3821 (76%)	1984 (39%)
<i>Hordeum vulgare</i>	23078	22:56:59	3.6 s	22145 (95%)	21564 (93%)	-

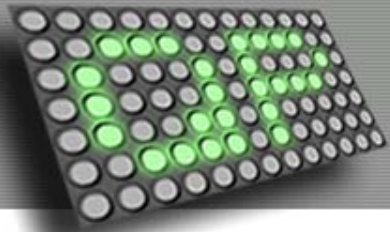
Primer pairs designed for hypothetical high-throughput experiments, for random transcripts of each species. The numbers of successfully designed primer pairs for the different specificity ranks and the search times are reported for each species (percentages refer to the total number of transcripts tested).

¹ Percentages indicate for how many of the transcripts primer pairs of at least the rank given were identified. ² 'Acceptable' amplifies only the specific sequence, but one primer has a high similarity with a non-target sequence and the primer pair might amplify genomic DNA. ³ 'Good' amplifies only the target sequence, but one primer has a high similarity with a non-target sequence or the pair might amplify genomic DNA. This is the highest possible rank for primer pairs designed for species without a genome annotation. ⁴ 'Very good' amplifies only the target sequence, both primers are highly specific to this sequence and will not amplify genomic DNA.



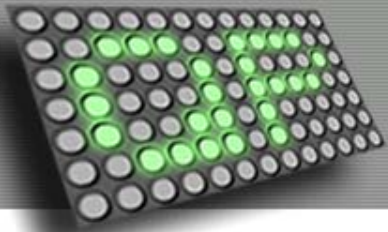
Public server usage

- Since Nov 1st 2008: 200 occasional users;
more than 100 frequent users
 - From all over the world
 - Working with all kinds of organisms, e.g. yeast,
tomato, water buffalo and human
- Sequence databases added on request



Summary

- QuantPrime is an easy-to-use yet flexible primer design tool for low- to high-throughput qPCR
- Robust; primer pairs have > 96 % success in experimental tests
- The public server, which is free to use for non-profit researchers, has many sequence sets installed, even for uncommon species not covered by commercial assays and software
- The link: <http://www.quantprime.de/>
 - Check out the paper for more details on algorithms and parameters (link on website)
- Coming up:
 - TF platform for tomato designed with QuantPrime, in testing
 - TF platform for *Physcomitrella* (moss) planned
 - RAM and CPU boost for the public server to better cope with big genomes
 - RDML export
 - More features...



Thanks to...

- The QuantPrime co-developers:



Dr. Mirosław
Kwaśniewski



Dr. Diego M.
Riaño-Pachón



Prof. Dr. Bernd
Müller-Röber

- The rest of AG MürRö at Uni Potsdam and MPIMP
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 - MK: DAAD, “Modern Applications of Biotechnology”, No. A/06/04209
 - DMRP: BMBF, GABI-FUTURE grant 0315046
 - BMR: BMBF, GoFORSYS, FKZ 0313924

- ... you for your attention!

